

## **BIOLOGICAL ANTAGONISTS FOR CONTROL OF PARASITIC DISEASES**

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**Abstract:** The present literature reviews biological methods for control of parasitic diseases in animals with a hope to encourage eminent scientists and practicing veterinarians in the field of veterinary practice to apply these methods as a component of integrated insect management.

**Keywords:** biological methods, biodegradable, Plant extracts, Fungi species, potential antagonists.

### **Introduction**

Parasitic diseases are a major constraint to efficient livestock production throughout many parts of the world (Morrison and Tomley, 2016). Chemical control methods have been used since years conventionally for controlling parasitic diseases. Largely because of the remarkable developments in these products in terms of efficacy, safety, spectrum of activity while remaining relatively inexpensive, livestock producers have relied almost exclusively on their use (Waller and Faedo *et al.*, 1993). Extensive usage of drugs has resulted in resistance in parasites' populations (Minho *et al.*, 2008). Moreover, the rapidly increasing number of organic farms in many countries over the last ten years and the consumer pressure for reduced chemical residues in products require finding of new, safe for the animals and environment, methods for control of parasitic diseases (Sanyal, 2009). Thus, biological control measures of animal parasites have been gaining importance in controlling parasitic population. The present literature reviews biological control methods of parasitic diseases which would be reasonably cheap and biodegradable.

### **1. Plant extracts:**

#### **(i) Condensed tannins**

Paolini *et al.* (2003) have assessed the possible impact of condensed tannins on goats infected with adult *Haemonchus contortus* and reported that major consequence of tannin consumption in goats reduced worm fecundity and egg output.

Paolini *et al.* (2003) investigated the effects of condensed tannins (CT) on adult populations of *Trichostrongylus colubriformis* and *Teladorsagia circumcincta* and reported that tannin administration was associated with a decrease in egg excretion and a decrease in female fecundity.

Minho *et al.* (2008) have investigated the anthelmintic effect of condensed tannin extracts (CTE) from *Acacia molissima* on lambs naturally infected with *H. contortus* and *T. colubriformis* and confirmed the anthelmintic effects of CTE on gastrointestinal nematodes in lambs and demonstrated the potential use of CTE as an alternative endoparasite control in livestock.

## (ii) Other plant products

Administration of plant-based preparation Loshtak *per os* as tablets made from standardized dust of bryony (*Bryonia alba*) roots was studied in experimental dictyocaulosis of lambs (Movsesyan *et al.*, 1994), in experimental ascaridosis of chickens (Chubarian *et al.*, 2002). It has been shown that the preparation increases the animals' natural resistance to these infections through activation of immunocompetent cells and phagocytes (Movsesyan *et al.*, 2004).

Hounzangbe–Adote *et al.*, (2005) studied the *in vitro* effects of extracts of four tropical plants (*Zanthoxylum zanthoxyloides*, *Newbouldia laevis*, *Morinda lucida* and *Carica papaya*) on the egg, infective larvae and adult worms of *Trichostrongylus colubriformis* and stated the presence of some anthelmintic properties associated with the four plants, which are traditionally used by small farmers in western Africa.

Hordegen *et al.*, (2006) investigated the effect of extracts of six different plant species [Bromelain, the enzyme complex of the stem of *Ananas comosus* (Bromeliaceae), the ethanolic extracts of seeds of *Azadirachta indica* (Meliaceae), *Caesalpinia crista* (Caesalpinaceae) and *Vernonia anthelmintica* (Asteraceae), and the ethanolic extracts of the whole plant of *Fumaria parviflora* (Papaveraceae) and of the fruit of *Embelia ribes* (Myrsinaceae)] against exsheathed infective larvae of *H. contortus* using a modified methyl-thiazolyl-tetrazolium (MTT) reduction assay and reported that these extracts had an anthelmintic efficacy of up to 93 %, relative to pyrantel tartrate.

## 2. Fungi species

Larsen *et al.* (1995) have investigated the effect of the *Duddingtonia flagrans* on calves infected by trichostrongyles under natural grazing conditions and reported that daily feeding with the microfungus *D. flagrans* during the first 2 months of the season reduced

herbage infectivity and lowered acquisition of *Ostertagia* sp. and *Cooperia* sp. faecal larval cultures revealed that the reduction in the infectivity was due to nematode-destroying effects of the fungi in the dung excreted by the fungus-treated calves (Salkova *et al.*, 2014).

Chandrawathani *et al.* (2003) conducted trials aimed at control of nematode *H. contortus* in small ruminants in a wet, tropical environment using *D. flagrans* and reported that at lower dosage (125 000 spores/kg live weight per day), larval population reduced to 80 and 90 % while at the higher dose rate (250 000 spores/kg live weight per day), there has been virtually complete suppression (>99 % reduction) of larval recovery.

*Beauveria bassiana* and *Metaregium anisopeliae* have been found to cause mortality in some ticks and insects (Yadav *et al.*, 2017). *M. Anisopheles* have been successfully tried against the *Boophilus microplus* and is giving encouraging results (Ment *et al.*, 2010).

### **3. Bacteria**

Strains of the entomopathogenic bacterium *Bacillus thuringiensis* are among the most widely used antagonists in biological control of insects where target insects are killed by a gut toxin which is released from crystal proteins in the bacterial spores (Pirali-Kheirabadi, 2012). In some countries, *B. thuringiensis* var. *israelensis* is available for control of mosquito larvae (Ravensberg 1994). The black fly *Simulium damnosum* which is an important vector of *Onchocerca volvulus*, causative agent of river blindness, is susceptible to *B. thuringiensis* var. *Israelensis* (Pirali-Kheirabadi, 2012).

### **4. Amphibians and fishes**

The water-tortoise *Pelomedusa subrufa* was reported to be able to remove ticks from black rhinos in a streambed (Mwangi *et al.*, 1991). The mosquito larvae may be controlled biologically by predatory fish such as *Gambusia affinis* and the Guppy *Poecilia* (Pirali-Kheirabadi, 2012).

### **5. Rodents and mammals**

*Sorex araneus* prey on ticks and at times preferred them to alternative foods. (Mwangi *et al.*, 1991). Shrews seem to locate hidden ticks by their smell while mice and rats are often cited as preying on ticks (Pirali-Kheirabadi, 2012).

### **6. Ants: (Hymenoptera) and beetles**

The predation in open areas was two to eight times higher than in woody ones. (Mwangi *et al.*, 1991). Fire ants predate on the *amblyomma* tick (Yadav *et al.*, 2017). Application of *S. invicta* in the United States markedly reduced the number of anaplasmosis in seropositive cattle in Louisiana (Jemal and Hugh-Jones., 1993). Wild rabbits living in

Formica polyctena infested plots had far fewer ticks than those in ant-free plots and rabbits sprayed with formic acid were free of ticks for at least 5 days (Buttner., 1987).

### Conclusion

Biological control of parasites though not succeeded well in Veterinary practice, it provides promising results as a substitute to chemical control and still further research is needed for the development of potential antagonists which can be affordable by a farmer.

### References

- [1] Chandrawathania P., Jamnaha O., Waller P.J. (2003). Biological control of nematode parasites of small ruminants in Malaysia using the nematophagous fungus *Duddingtonia flagrans*. Vet. Parasitol., 117(3):173–183
- [2] Chubarian F.A., Petrosyan R.A., Movsesyan S.O., Boyakhcyan G.A. (2002). Effects of Loshtak on natural resistance of chickens at experimental *Ascaridia galli* infection. Konf. Theory and practice of fighting parasitic diseases., 3: 374–375.
- [3] De S., Sanyal P.K. (2009). Biological Control of Helminth Parasites by Predatory Fungi. VetScan. 4, (1): 1–8
- [4] Hordegen P., Cabaret J., Hertzberg H. (2006). *In vitro* screening of six anthelmintic plant products against larval *Haemonchus contortus* with a modified methyl-thiazolyl-tetrazolium reduction assay. J. of Ethnopharmacol., 108 (1): 85–89.
- [5] Hounzangbe–Adote S., Fouraste I., Moutairou K., Hoste H. (2005). *In vitro* effects of four tropical plants on the activity and development of the parasitic nematode, *Trichostrongylus colubriformis*. J. of Helminthol. 79(01):29–33.
- [6] Ivan Morrison and Fiona Tomley, Development of vaccines for parasitic diseases of animals: Challenges and opportunities. Parasite immunology, DOI: 10.1111/pim.12398.
- [7] Larsen M., Nansen P., Wolstrup J. (1995). Biological control of trichostrongyles in calves by the fungus *Duddingtonia flagrans* fed to animals under natural grazing conditions. Vet. Parasitol. 60 (3): 321–330.
- [8] Ment D, Gindin G, Rot A, Soroker V, Glazer I, Bare S. (2010). Novel Technique for Quantifying Adhesion of *Metarhizium anisopliae* Conidia to the Tick Cuticle. Applied and Environmental Microbiology. 2010; 3521- 3528.
- [9] Minho A.P., Bueno I.C.S., H. Louvandini H. (2008). Effect of *Acacia molissima* tannin extract on the control of gastrointestinal parasites in sheep. Animal Feed Sci. and Technol., 147(1–3): 172–181.

- [10] Minho A.P., Bueno I.C.S., H. Louvandini H. (2008). Effect of *Acacia molissima* tannin extract on the control of gastrointestinal parasites in sheep. *Animal Feed Sci. and Technol.* 147(1): 172–181.
- [11] Movsesyan S.O., Boyakhchyan G.A., Chubaryan F.A. (1994). Effects of plant preparation Loshtak on immune status of lambs in case of experimental dictyocaulosis. In: *Differential diagnostics of parasites and aspects of parasite-host relationships.* – Moscow: Institute of Parasitology, P. 13–19.
- [12] Movsesyan S.O., Chubaryan F.A., Petrosyan R.A. (2004). Animal non-specific resistance and ways of its correction in some helminthoses. *Transac. of the Institute of Parasitol. Progress in General Parasitology. Moscow*, 44:221–230.
- [13] Mwangi EN, Dipeolu OO, Newson RM, Kaaya GP and Hassan SM. 1991. Predators, parasitoids and pathogens of ticks: a review. *Biocont. Sci. Tech.* 1:147-156.
- [14] Paolini V., Frayssines A., De La Farge F. (2003). Effects of condensed tannins on established populations and on incoming larvae of *Trichostrongylus colubriformis* and *Teladorsagia circumcincta* in goats. *Vet. Res.* 34: 331–339.
- [15] Pirali-Kheirabadi, K. (2012). Biological Control of Parasites. In *Parasitology. InTech*. P. 53-66.
- [16] Ravensberg WJ. 1994. Biological control of pests: current trends and future prospects. Brighton Crop Protection Conference-Pest and Diseases. British Crop Protection Council, Thornton Heath, UK, pp. 591-600.
- [17] Salkova, D., Panayotova–Pencheva, M., Movsesyan, S., Be'er, S., & Voronin, M. (2014). Alternative methods for control of parasitic diseases in animals. *РОССИЙСКИЙ ПАРАЗИТОЛОГИЧЕСКИЙ*, 93.
- [18] Waller PJ: Sustainable nematode parasite control strategies for ruminant livestock by grazing management and biological control. *Animal Feed Science and Technology* 2006, 126:277-289.
- [19] Yadav, P. K., Rafiqi, S. I., Panigrahi, P. N., Kumar, D., Kumar, R., & Kumar, S. (2017). Recent trends in control of ectoparasites: *A.* 5(2): 808-813.